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Solvent Hold Tank Sample Results for MCU-15-750-751-752-: June Monthly Sample

F. F. Fondeur
K. M. L. Taylor-Pashow
October 2015
SRNL-STI-2015-00469, Revision 0
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F. F. Fondeur
K. M. L. Taylor-Pashow

October 2015
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ACKNOWLEDGEMENTS

We acknowledged and appreciated Mr. Daniel Jones for the titration of the solvent.
EXECUTIVE SUMMARY

Savannah River National Laboratory (SRNL) received one set of Solvent Hold Tank (SHT) samples (MCU-15-750, MCU-15-751, and MCU-15-752), pulled on 06/22/2015 for analysis. The samples were combined and analyzed for composition. Analysis of the composite sample MCU-15-750-751-752 indicated a low concentration (~ 49 % of nominal) of the suppressor (TiDG) and slightly lower than nominal concentrations of the extractant (MaxCalix), and of the modifier (Cs-7SB) in the solvent. This analysis confirms the addition of TiDG, MaxCalix, and modifier to the solvent in February 2015. Based on the current monthly sample, the levels of TiDG, MaxCalix, and modifier are sufficient for continuing operation without adding a trim at this time but it is recommended that an addition of TiDG, modifier and Isopar™L should be made in the near future.

No impurities above the 1000 ppm level were found in this solvent by the Semi-Volatile Organic Analysis (SVOA). No impurities were observed in the Hydrogen Nuclear Magnetic Resonance (HNMR). In addition, up to 13.9 micrograms of mercury per gram of solvent (or 11.5 µg/mL) was detected in this sample.

The laboratory will continue to monitor the quality of the solvent in particular for any new impurities or degradation of the solvent components.
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<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>BOBCalixC6</td>
<td>Calix[4]arene-bis(tert-octylbenzo-crown-6)</td>
</tr>
<tr>
<td>FT-HNMR</td>
<td>Fourier Transform Hydrogen Nuclear Magnetic Resonance</td>
</tr>
<tr>
<td>FTIR</td>
<td>Fourier transform infra-red spectroscopy</td>
</tr>
<tr>
<td>HPLC</td>
<td>High Performance Liquid Chromatography</td>
</tr>
<tr>
<td>ISDP</td>
<td>Integrated Salt Disposition Project</td>
</tr>
<tr>
<td>MCU</td>
<td>Modular Caustic-Side Solvent Extraction Unit</td>
</tr>
<tr>
<td>MaxCalix</td>
<td>1,3-alt-25,27-Bis(3,7-dimethyloctyloxy)calix[4]arene-benzocrown-6</td>
</tr>
<tr>
<td>NGS</td>
<td>Next Generation Solvent</td>
</tr>
<tr>
<td>RSD</td>
<td>Relative Standard Deviation or the absolute value of the Coefficient of Variation</td>
</tr>
<tr>
<td>SHT</td>
<td>Solvent Hold Tank</td>
</tr>
<tr>
<td>SRNL</td>
<td>Savannah River National Laboratory</td>
</tr>
<tr>
<td>SVOA</td>
<td>Semi-Volatile Organic Analysis</td>
</tr>
<tr>
<td>TiDG</td>
<td>$N,N',N''$–tris(3,7-dimethyloctyl)guanidine</td>
</tr>
<tr>
<td>TOA</td>
<td>Trioctylamine</td>
</tr>
<tr>
<td>XRF</td>
<td>X-Rays Fluorescence</td>
</tr>
</tbody>
</table>
1.0 Introduction

In late FY13, the Modular Caustic-Side Solvent Extraction Unit (MCU) switched to the Next Generation Solvent (NGS) flow sheet. Facility personnel implemented the switch by adding a non-radioactive, NGS “cocktail” containing the new extractant (MaxCalix) and a new suppressor (TiDG) to the SHT heel. The resulting “blend” solvent (“NGS Blend solvent”) is essentially NGS with residual amounts of BOBCalixC6 and trioctylamine (TOA). SHT samples are sent to SRNL to examine solvent composition changes over time. On June 22, 2015, Operations personnel pulled and delivered three samples from the SHT (MCU-15-750, MCU-15-751, and MCU-15-752) for analysis. These samples are intended to verify that the solvent is within the specified composition range. A baseline “scratch” solvent (a scratch solvent is a preparation of all 6 solvent components at the same time to generate a solution of the appropriate composition that approximates the blend of cocktail and heel solvent) was prepared in the lab (May 14, 2014) and used for comparison and evaluation. The results from the analyses are presented in this document.

2.0 Experimental Procedure

2.1 Experimental Procedure

A summary of relevant and recent trims to the MCU solvent as well as the arrival date of the samples currently being studied are shown in Table 2-1. On February 22, 2015, a trim addition was made to MCU that was 2.23E4 grams of modifier, 838 grams of TiDG, and 2.23E3 grams of MaxCalix in 10 gallons of Isopar™L.

<table>
<thead>
<tr>
<th>Event</th>
<th>Date</th>
</tr>
</thead>
<tbody>
<tr>
<td>February solvent trim added to MCU</td>
<td>February 22, 2015</td>
</tr>
<tr>
<td>20 gallons of Isopar™L added to MCU</td>
<td>March 6, 2015</td>
</tr>
<tr>
<td>SHT sample MCU-15-661-662-663</td>
<td>April 2, 2015</td>
</tr>
<tr>
<td>10 gallons of Isopar™L added to MCU</td>
<td>May 6, 2015</td>
</tr>
</tbody>
</table>

Samples shown in Table 2-1 were received in p-nut vials containing ~10 mL each (see Fig 1). Once taken into a radioactive hood, the samples were visually inspected and analyzed for pH. MCU-15-750, MCU-15-751, and MCU-15-752 were composited before use. Aliquots of the composited sample were removed to perform the following analysis: Density, semi-volatile organic analysis (SVOA), high performance liquid chromatography (HPLC), titration, gamma counting and Fourier-Transformed Hydrogen Nuclear Magnetic Resonance (FT-HNMR). Fourier-Transformed Infrared Spectroscopy was not performed as the portion of the sample set aside for this measurement was accidentally kept in a Teflon bottle with a large overhead space that resulted in significant Isopar™L evaporation. Results from analytical measurements were compared with the theoretical values shown in Table 2-2.
Table 2-2 Nominal concentrations of the relevant components in NGS Blend

<table>
<thead>
<tr>
<th>Component</th>
<th>mg/L</th>
<th>Molar</th>
</tr>
</thead>
<tbody>
<tr>
<td>MaxCalix</td>
<td>~44,400</td>
<td>~0.0465</td>
</tr>
<tr>
<td>BOBCalixC6*</td>
<td>&lt;4,030</td>
<td>&lt;0.0035</td>
</tr>
<tr>
<td>TOA*</td>
<td>&lt;530</td>
<td>&lt;0.0015</td>
</tr>
<tr>
<td>Modifier</td>
<td>~169,000</td>
<td>~0.50</td>
</tr>
<tr>
<td>TiDG</td>
<td>~1440</td>
<td>~0.003</td>
</tr>
<tr>
<td>Isopar™L</td>
<td>~623,000</td>
<td>~74 wt%</td>
</tr>
</tbody>
</table>

*Values represent starting values when NGS blend was implemented. These components are no longer added to or refurbished in MCU.

2.2 Quality Assurance

Requirements for performing reviews of technical reports and the extent of review are established in manual E7 2.60. SRNL documents the extent and type of review using the SRNL Technical Report Design Checklist contained in WSRC-IM-2002-00011, Rev. 2.

3.0 Results and Discussion

The p-nut vials from MCU-15-750, MCU-15-751, and MCU-15-752 were examined and found to contain a single phase liquid with no apparent solids contamination or cloudiness. All samples had a pH value of 5.5. No unusual reactions, solids, foaming, or immiscible layers were observed after combining the samples into one (MCU-15-750-751-752). Table 3-1 contains the results for the MCU-15-750-751-752 sample.

*Isopar™L and Modifier Levels*

A density measurement of the sample gave a result of 0.830 g/mL (0.09% RSD) (or 0.828 g/mL at 25 °C when corrected for temperature using the CSSX temperature correction formula) for MCU-15-750-751-752 at 23 °C. The calculated density (0.828 g/mL) for MCU-15-750-751-752 is 1% below the calculated density for the standard sample (0.835 g/mL at 25 °C for the scratch blend made in the laboratory)\(^1\). Using the density as a starting point, we know that the concentration level of the Isopar™L component in the sample should be equal to or slightly above the nominal value.

---

\(^1\) A second standard was prepared on December 12, 2014
An examination of Table 3-1 shows that the Isopar™ L concentration is at its nominal value while the modifier concentration is correspondingly slightly lower (9% lower) than its nominal value. Of all the methods listed, density has the lowest uncertainty. Thus, the final reported values are closer to the density measurement. The last Isopar™L trim addition to MCU was on May 6, 2015.

All measurements indicate the Isopar™ L level is at nominal value while the modifier concentration level is below its nominal value (see Fig. 2). The modifier level appears to trend up and down possibly reflecting randomness in the process of mixing, sampling and analyzing it. The relatively lower modifier concentration explains why the measured density is slightly below the standard sample density. The accuracies of the different measurements were within expectation as reflected in the total mass sum of the “average” results listed in Table 3-1. They added up to 0.829 ± 0.020 g/mL. Their sum is consistent with the measured and temperature corrected (to 25 °C) value of 0.828 g/mL, but it is below the measured and corrected to 25 °C mass concentration (density) of the standard (0.835 g/mL).

Figure 1. Typical appearance of the three vials MCU-15-750, MCU-15-751, and MCU-15-752
### Table 3-1 Sample Results for MCU-15-750-751-752

<table>
<thead>
<tr>
<th>Analysis</th>
<th>Method^6</th>
<th>LIMS #</th>
<th>Result (mg/L)^8</th>
<th>Nominal* Result (mg/L)</th>
<th>% of (Result ÷ Nominal Result)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Isopar® L</td>
<td>FT-HNMR</td>
<td>NA</td>
<td>6.37E+05</td>
<td>6.23E+05</td>
<td>102</td>
</tr>
<tr>
<td>Isopar® L</td>
<td>Density</td>
<td>NA</td>
<td>6.25E+05</td>
<td>6.23E+05</td>
<td>100</td>
</tr>
<tr>
<td>Average^7</td>
<td>All</td>
<td>NA</td>
<td>6.25E+05</td>
<td>6.23E+05</td>
<td>100</td>
</tr>
<tr>
<td>Modifier</td>
<td>HPLC</td>
<td>300318159</td>
<td>1.61E+05</td>
<td>1.69E+05</td>
<td>93</td>
</tr>
<tr>
<td>Modifier</td>
<td>FT-HNMR</td>
<td>NA</td>
<td>1.57E+05</td>
<td>1.69E+05</td>
<td>92</td>
</tr>
<tr>
<td>Modifier</td>
<td>Density</td>
<td>NA</td>
<td>1.53E+05</td>
<td>1.69E+05</td>
<td>91</td>
</tr>
<tr>
<td>Average^7</td>
<td>All</td>
<td>NA</td>
<td>1.54E+05</td>
<td>1.69E+05</td>
<td>91</td>
</tr>
<tr>
<td>TiDG</td>
<td>Titration</td>
<td>NA</td>
<td>6.85E+02</td>
<td>1.44E+03</td>
<td>48</td>
</tr>
<tr>
<td>TiDG</td>
<td>FT-HNMR</td>
<td>NA</td>
<td>8.02E+02</td>
<td>1.44E+03</td>
<td>56</td>
</tr>
<tr>
<td>Average^6</td>
<td>All</td>
<td>NA</td>
<td>7.03E+02</td>
<td>1.44E+03</td>
<td>49</td>
</tr>
<tr>
<td>trioctylamine</td>
<td>Titration</td>
<td>NA</td>
<td>3.21E+02</td>
<td>5.30E+02</td>
<td>61</td>
</tr>
<tr>
<td>Average^6</td>
<td>All</td>
<td>NA</td>
<td>3.21E+02</td>
<td>5.30E+02</td>
<td>61</td>
</tr>
<tr>
<td>MaxCalix</td>
<td>FT-HNMR</td>
<td>NA</td>
<td>4.53E+04</td>
<td>4.44E+04</td>
<td>102</td>
</tr>
<tr>
<td>MaxCalix</td>
<td>HPLC</td>
<td>300318159</td>
<td>4.53E+04</td>
<td>4.44E+04</td>
<td>102</td>
</tr>
<tr>
<td>Average^6</td>
<td>All</td>
<td>NA</td>
<td>4.53E+04</td>
<td>4.44E+04</td>
<td>102</td>
</tr>
<tr>
<td>BOBCalixC6</td>
<td>HPLC</td>
<td>300318159</td>
<td>3.00E+03</td>
<td>4.03E+03</td>
<td>74</td>
</tr>
<tr>
<td>Average^6</td>
<td>All</td>
<td>NA</td>
<td>3.00E+03</td>
<td>4.03E+03</td>
<td>74</td>
</tr>
<tr>
<td>Density (g/mL)</td>
<td>Direct Measurement</td>
<td>NA</td>
<td>0.8281</td>
<td>0.835</td>
<td>99</td>
</tr>
</tbody>
</table>

^6 Analytical uncertainty is 20% for SVOA and 10% for HPLC. Titration method uncertainty is 10% for TiDG and 20% for TOA. Density results from the average of replicate volumetric trials typically have a percentage standard deviation of <3% between each value and the average. NMR analytical uncertainty is 10% for the modifier and 13% for MaxCalix, 14% for Isopar™ L, and 20% for TiDG. N/A = Not Applicable. Density estimations assume the combined weight percent of TiDG, MaxCalix, BOBCalix, and TOA to be approximately 6%.

^* Nominal value is the expected value for freshly prepared blended solvent with a target density of 0.8352 g/mL at 25 °C.

^8 x = \frac{\sum (x_i - \bar{x})^2}{\sum (x_i - \bar{x})}; \bar{x} \text{ stands for the concentration obtained at a given method and } \sigma \text{ is the corresponding uncertainty.}

^6 FTIR quantification was not conducted on this sample since the sample was unintentionally kept in a large overhead bottle where Isopar™ L evaporated into.
Suppressors Levels

The average TiDG concentration level (7.03 ± 0.7E2 mg/L) is at 49% of its nominal value of 1440 mg/L despite the trim addition in February 2015. The suppressor concentration is above the minimum recommended operating level 479 mg/L and thus, the solvent does not require a TiDG addition at the time sample MCU-15-750-751-752 was collected. Inferring from past TiDG concentrations level trends and in the absence of new additions or new removal mechanisms, the TiDG concentration is expected to drop as shown in Fig. 3. The TOA concentration appears to have risen to 321 ± 64 mg/L (in the previous month the TOA level was 290 mg/L). These numbers are within analytical error. In addition, looking at the slope of the TOA concentration data, it appears to trend upwards with time. The upward trend appears to correlate with the disappearance of TiDG (or the disappearance of Isopar™ L) concentration. This correlation is believed to be due to the titration of TiDG’s decomposition products and to the evaporation of Isopar™ L. Since MCU no longer adds TOA, a drop in TOA concentration is expected. However, a closer inspection of Fig. 2 reveals the TOA level increasing in the last three MCU samples. The rise is perhaps due to TiDG degradation into primary amines, which have previously been identified as degradation products of the suppressor when heated. The primary amine degradation products would likely have a similar pKa to the TOA (tertiary amine) making the equivalent points coincide.
**Extractant Levels**

The average MaxCalix level is 4.53 E4 mg/L (±13%) which is 2% above the nominal concentration and it is within the 95% confidence level of the analytical measurement as shown in Figure 4. The residual concentration of BOBCalix level is currently at 75% of the level measured when the NGS was implemented in late FY13.

**Gamma Level**

The gamma measurement of MCU-15-750-751-752 is 7.98E+04 dpm/mL (±5%). This level of activity, which is two orders of magnitude lower than observed in sample MCU-15-710-711-712, is consistent with previous levels or baseline level (see Fig. 5) where the solvent performed (extraction and stripping) as expected.

**Impurities**

No impurities were seen at the 1000 ppm level or higher as indicated by the SVOA method. No impurities were observed in the FTIR spectrum of this sample.

A few mL of the blended solvent were digested and analyzed for mercury (by the cold vapor method and the XRF method). The total mercury level in the solvent measured 13.9 µg/g_{solvent} (20% st.dev.) or 11.5 µg/mL_{solvent}. This is significantly higher than the solubility of metallic Hg in dodecane (~3 ppm) implying that other solubility-enhancing mechanisms are at play (for example extraction or sorption or trapped solids) or a more soluble form of mercury is present (organo-mercury like ethyl or dimethyl mercury). Organo-mercury compounds were recently detected in Tank 22H. For 200 gallons of solvent (757.1 L) and assuming a density of 0.828 g/mL, the solvent could contain a total of 8.7 ± 2 g of mercury. See figure 6 for a comparison of recent SHT mercury concentrations.

**Recommendation**

The current analysis indicates low modifier and TiDG levels in this solvent relative to the standard. The TiDG level is trending downwards similarly to previous trends observed after a TiDG trim addition is made to the solvent. However, the current TiDG level is below the minimum operating recommended level (958 mg/L_{solvent}) but above the minimum recommended level (479 mg/L_{solvent}). There is sufficient TiDG in the solvent for continuing operation without adding a trim until the next monthly sample. However, to remain two-sigma above the minimum recommended level, it is recommended that TiDG, modifier and appropriate amounts of Isopar™L be added to restore the components to nominal levels. Similarly, the modifier level (1.54E5 mg/L) is well above the minimum modifier level at which the MaxCalix becomes insoluble in the solvent.
Figure 3. Suppressor concentration as measured by titration in SHT samples since NGS implementation. The minimum recommended is 479 mg/L for TiDG.

Figure 4. MaxCalix concentration as measured by HPLC and FT-HNMR of recent samples since NGS implementation (44,400 mg/L is the nominal concentration).
Figure 5. The gamma count of selected SHT samples. One standard deviation is 5%.
Figure 6. Total mercury in recent SHT samples. One standard deviation is 20%.

4.0 Conclusions

SRNL received one set of SHT samples (MCU-15-750, MCU-15-751, and MCU-15-752), pulled on 06/22/2015 for analysis. The samples were combined and analyzed for composition. Analysis of the composite sample MCU-15-750-751-752 indicated a low concentration (~49 % of nominal) of the suppressor (TiDG) and concentrations of the extractant (MaxCalix), and of the modifier (CS-7SB) in the solvent that were slightly lower than nominal. This analysis confirms the addition of TiDG, MaxCalix, and modifier to the solvent in February 2015. Based on the current monthly sample, the levels of TiDG, MaxCalix, and modifier are sufficient for continuing operation without adding a trim at this time but it is recommended that an addition of TiDG, modifier and Isopar™L should be made in the near future.

No impurities above the 1000 ppm level were found in this solvent by the Semi-Volatile Organic Analysis (SVOA). No impurities were observed in the Hydrogen Nuclear Magnetic Resonance (HNMR). However, up to 13.9 ± 3 micrograms of mercury per gram of solvent (or 11.54 µg/mL) was detected in this sample.

The laboratory will continue to monitor the quality of the solvent in particular for any new impurities or degradation of the solvent components.
5.0 References

1 W. M. Matthews, HLW-CRF-10006, Rev. 0, May 18, 2010.


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