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# Solvent Hold Tank Sample Results for MCU-14-1172-1173: December 2014 Monthly Sample

F. F. Fondeur

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February 2015

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## **EXECUTIVE SUMMARY**

SRNL received one set of SHT samples (MCU-14-1172-1173 pulled on 12/30/2014) for analysis. The samples were combined and analyzed for composition. Analysis of the composite sample MCU-14-1172-1173 indicated low concentrations of the suppressor (TiDG), of the extractant (MaxCalix), and of the modifier (CS-7SB) in the solvent relative to their nominal values. This analysis confirms a downward trend of these components. No impurities were found in this solvent. The laboratory will continue to monitor the quality of the solvent in particular for any new impurity or degradation of the solvent components.

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## LIST OF ABBREVIATIONS

BOBCalixC6	Calix[4]arene-bis( <i>tert</i> -octylbenzo-crown-6)
FT-HNMR	Fourier Transform Hydrogen Nuclear Magnetic Resonance
FTIR	Fourier transform infra-red spectroscopy
HPLC	High Performance Liquid Chromatography
ISDP	Integrated Salt Disposition Project
MCU	Modular Caustic-Side Solvent Extraction Unit
MaxCalix	1,3- <i>alt</i> -25,27-Bis(3,7-dimethyloctyloxy)calix[4]arene-benzocrown-6
NGS	Next Generation Solvent
RSD	Relative Standard Deviation or the absolute value of the Coefficient of Variation
SHT	Solvent Hold Tank
SRNL	Savannah River National Laboratory
SVOA	Semi-Volatile Organic Analysis
TiDG	<i>N,N',N''</i> -tris(3,7-dimethyloctyl)guanidine
TOA	Trioctylamine

## 1.0 Introduction

In late FY13, MCU switched to the Next Generation Solvent (NGS) flow sheet. Facility personnel implemented the switch by adding a non-radioactive, NGS “cocktail” containing the new extractant (MaxCalix) and a new suppressor (TiDG) to the SHT heel. The resulting “blend” solvent (“NGS Blend solvent”) is essentially NGS with residual amounts of BOBCalixC6 and trioctylamine (TOA). Solvent Hold Tank (SHT) samples are sent to Savannah River National Laboratory (SRNL) to examine solvent composition changes over time.<sup>1</sup> On December 30, 2014, Operations personnel delivered two samples from the SHT (MCU-14-1172 and MCU-14-1173) for analysis. These samples were intended to verify that the solvent is within the specified composition range. A baseline “scratch” solvent (a from-scratch solvent is a preparation of all 6 solvent components at the same time to generate a solution of the appropriate composition that approximates the blend of cocktail<sup>2</sup> and heel solvent) was prepared in the lab (November 29, 2014) and used for comparison and evaluation. The results from the analyses are presented in this document.

## 2.0 Experimental Procedure

### 2.1 Experimental Procedure

A summary of relevant and recent trims to the MCU solvent as well as the arrival date of the samples currently studied is shown in Table 2-1.

**Table 2-1. Log of recent trims to the MCU solvent and sample arrivals to SRNL**

Event	Date
22 gallons of Isopar <sup>TM</sup> L trim added to MCU	November 25, 2014
17 gallons of Isopar <sup>TM</sup> L trim added to MCU	December 19, 2014
SHT sample MCU-14-1172-1173	December 30, 2014

Samples shown in Table 2-1 were received in p-nut vials containing ~10 mL each (see Fig 1). Once taken into a radioactive hood, the samples were visually inspected and analyzed for pH. MCU-14-1172 and MCU-14-1173 were composited before use. Aliquots of the composited sample were removed to perform analysis by density, semi-volatile organic analysis (SVOA), high performance liquid chromatography (HPLC), titration, gamma counting, and Fourier-Transform Hydrogen Nuclear Magnetic Resonance (FT-HNMR). The Fourier-Transform Infra-Red spectroscopy (FTIR) instrument was not available at the time. Results from analytical measurements were compared with the theoretical values shown in Table 2-2.

**Table 2-2. Nominal concentrations of the relevant components in NGS Blend<sup>2</sup>**

Component	mg/L	Molar
MaxCalix	~ 44,400	~ 0.0465
BOBCalixC6*	< 4,030	< 0.0035
TOA*	< 530	< 0.0015
Modifier	~ 169,000	~ 0.50
TiDG	~1440	~ 0.003
Isopar <sup>TM</sup> L	~ 623,000	~ 74 wt%

\*Values represent starting values when NGS blend was implemented. These components are no longer added to or refurbish in MCU.

## 2.2 Quality Assurance

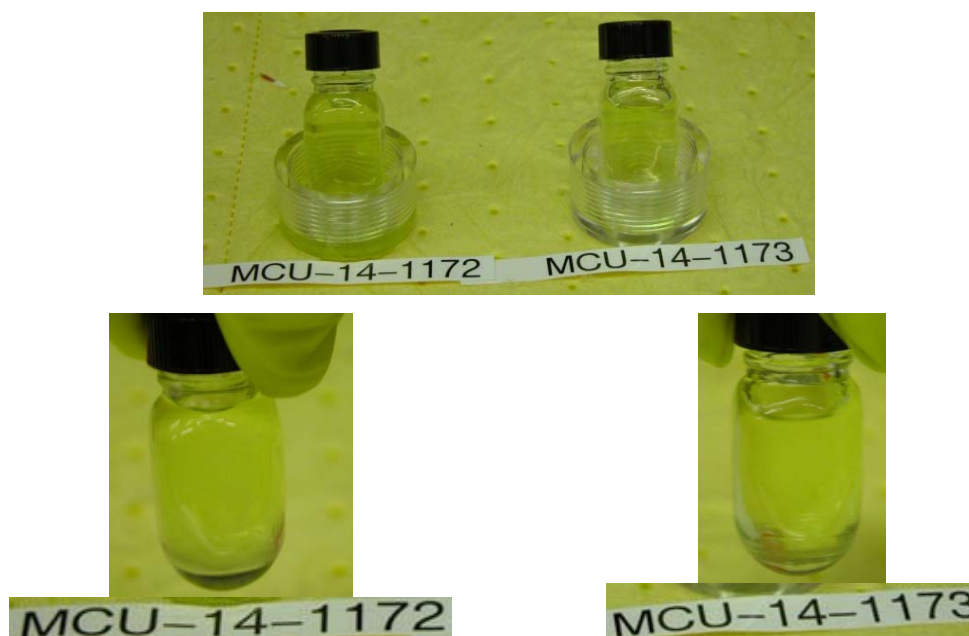
Requirements for performing reviews of technical reports and the extent of review are established in manual E7 2.60. SRNL documents the extent and type of review using the SRNL Technical Report Design Checklist contained in WSRC-IM-2002-00011, Rev. 2.

## 3.0 Results and Discussion

The p-nut vials from MCU-14-1172 and MCU-14-1173 were examined and found to contain a single phase liquid with no apparent solids contamination or cloudiness. All samples had a pH value of 5.5. No unusual reactions, solids, foaming, or immiscible layers were observed after combining the samples into one (MCU-14-1172-1173). Table 3-2 contains the results for the MCU-14-1172-1173 sample.

### *Isopar™ L and Modifier Levels*

Density measurement of the sample gave a result of 0.824 g/mL (0.1% RSD) (or 0.821g/mL at 25 °C when corrected for temperature using the CSSX temperature correction formula) for MCU-14-1172-1173 at 21.5 °C. The calculated density (0.821 g/mL) for MCU-14-1172-1173 is 1.5% below the calculated density for the standard sample (0.835 g/mL at 25 °C for the scratch blend made in the laboratory)<sup>1</sup>. Using the density as a starting point, we know that the concentration level of the Isopar™L component in the sample should be higher than its nominal value.



**Figure 1. Typical appearance of the two vials MCU-14-1172-1173**

An examination of Table 3-2 shows that the Isopar™ L concentration is higher than its nominal value while the modifier concentration is correspondingly lower than its nominal value. The spectroscopy method (FT-HNMR) reported higher values than the density and chromatography methods. Of all the methods listed, density has the lowest uncertainty. Thus, the final reported values are closer to the density measurement. The last Isopar™ L trim addition to MCU was on December 19, 2014. All measurements indicate the Isopar™ L level is above its nominal value while the modifier concentration level is below its

<sup>1</sup> A second standard was prepared on September 29, 2014

nominal value. The accuracy of the different measurements were within expectation as reflected in the total mass sum of the “average” results listed in Table 3-2, which add up to  $0.818 \pm 0.019$  g/mL. This is consistent with the measured and temperature corrected (to 25 °C) value of 0.820 g/mL, but it is below the measured and corrected to 25 °C mass concentration (density) of the standard (0.833 g/mL). Natural evaporation of the Isopar™ L may increase the concentration of these components.

#### *Suppressors Levels*

The TiDG concentration level (724 mg/L) is at 50 % of its nominal value of 1440 mg/L. This data point confirms the persistent downward trend in the TiDG concentration as shown in Fig. 2. The TOA concentration rose slightly to 348 mg/L (in the previous month the TOA level was 260 mg/L). Since MCU no longer adds TOA, the reason for this increase is unknown at this time. However, a simple correlation analysis between TiDG as determined by titration and HNMR and TOA as determined by titration appears to indicate a strong negative correlation between TiDG and TOA as determined by titration. The correlation is perhaps due to TiDG degradation to form primary amines, which have previously been identified as degradation products of the suppressor when heated.<sup>3</sup> The primary amine degradation products would likely have a similar pKa to the TOA (tertiary amine) making the equivalence points coincide.<sup>4</sup> Prior to the December sample analysis, there was a strong positive correlation (0.78) between TiDG and TOA based on titration analysis (both components decreased with time). After the correlation has become strongly negative (see Table 3-1)

**Table 3-1. Correlation coefficient between TiDG and TOA as determined by Titration and HNMR since the NGS-CSSX implementation. HNMR does not adequately measure TOA.**

	<i>TiDG titration</i>	<i>TOA titration</i>	<i>TiDG HNMR</i>
TiDG Titration	1		
TOA Titration	<b>-0.78</b>	1	
TiDG FT-HNMR	0.76	-0.63	1

#### *Extractant Levels*

The MaxCalix concentration continues its downward trend (4.6E4 mg/L) as shown in Fig. 3. The trend appears to be linear with time or volume of processed waste. The BobCalixC6 concentration slightly declined to 2,700 mg/L level possibly due to a dilution effect from the high Isopar™ L concentration. MCU no longer adds BOBCalixC6 to the solvent.

#### *Gamma Level*

The gamma measurement of MCU-14-1172-1173 is shown in Fig. 4. As can be seen in Fig. 4, the measured gamma level (2.95 E4 dpm/mL, ADS: 300315527) is consistent with previous gamma levels measurements of the solvent when there were no known processing issues at MCU. This indicates the solvent is stripping as intended.

#### *Impurities*

No impurities were detected by the SVOA method (ADS # 300315527) above the 1000 ppm level. No significant impurities were observed in the H-NMR spectrum of these samples.

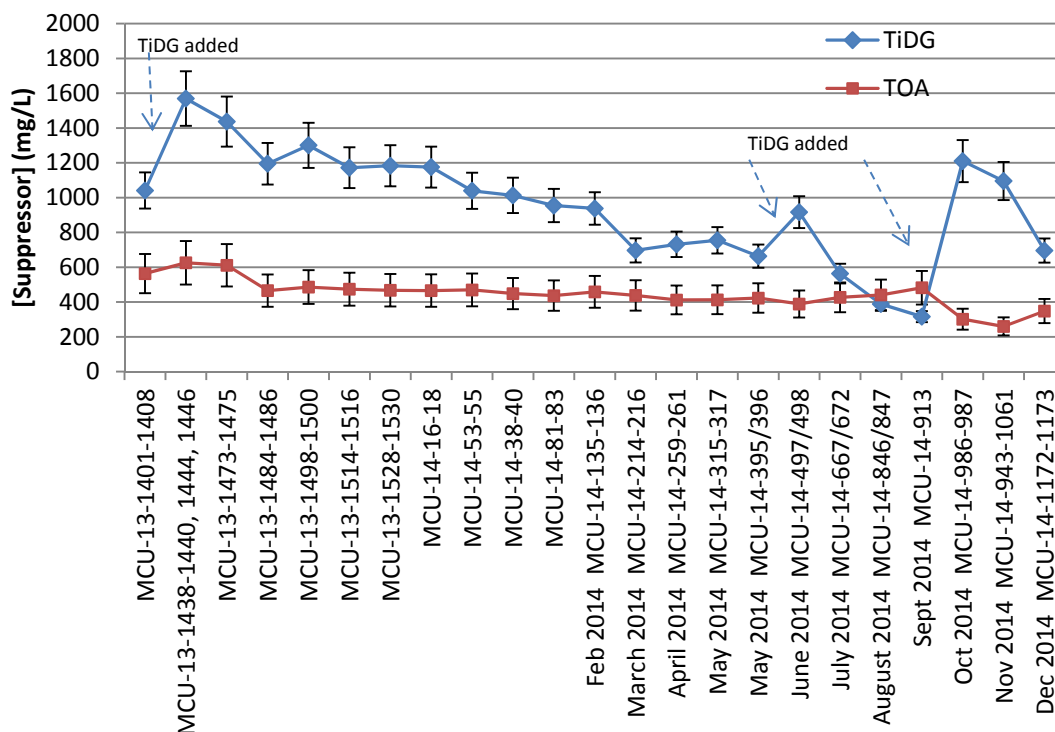
**Table 3-2. Sample Results for MCU-14-1172-1173**

Analysis	Method	LIMS #	Result (mg/L) <sup>#</sup>	Nominal* Result (mg/L)	% of (Result ÷ Nominal Result)
Isopar™ L	FT-HNMR	NA	6.51E+05	6.23E+05	104
Isopar™ L	Density*	NA	6.34E+05	6.23E+05	102
Average <sup>§</sup>	All	NA	6.35E+05	6.23E+05	102
Modifier	HPLC	300315527	1.40E+05	1.69E+05	83
Modifier	FT-HNMR	NA	1.45E+05	1.69E+05	86
Modifier	Density*	NA	1.37E+05	1.69E+05	81
Average <sup>§</sup>	All	NA	1.38E+05	1.69E+05	82
TiDG	Titration	NA	6.96E+02	1.44E+03	48
TiDG	FT-HNMR	NA	9.25E+02	1.44E+03	64
Average <sup>§</sup>	All	NA	7.24E+02	1.44E+03	50
trioctylamine	Titration	NA	3.48E+02	5.30E+02	66
Average <sup>§</sup>	All	NA	3.48E+02	5.30E+02	66
MaxCalix	FT-HNMR	NA	4.62E+04	4.44E+04	104
MaxCalix	HPLC	300315527	3.91E+04	4.44E+04	88
Average <sup>§</sup>	All	NA	4.12E+04	4.44E+04	93
BOBCalixC6	HPLC	300315527	2.97E+03	4.03E+03	74
Average <sup>§</sup>	All	NA	2.97E+03	4.03E+03	74
Density (g/mL)	Direct Measurement	NA	0.824	0.835	99

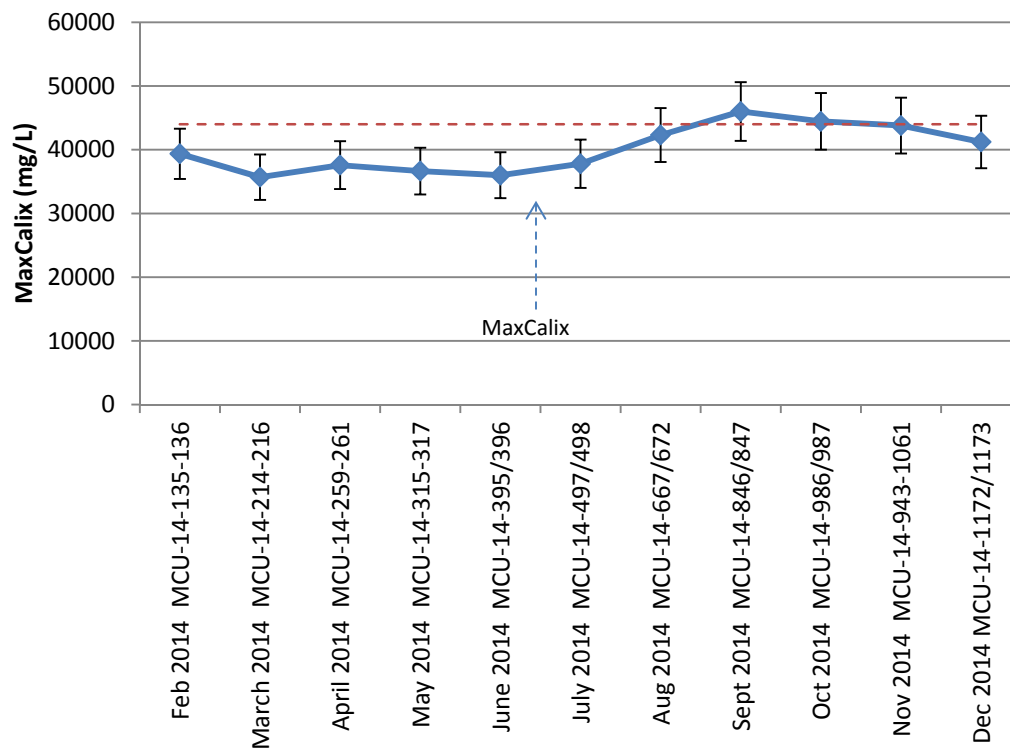
<sup>#</sup> Analytical uncertainty is 10% for HPLC. Titration method uncertainty is 10% for TiDG and 16% for TOA. Density results from the average of replicate volumetric trials typically have a percentage standard deviation of <3% between each value and the average. NMR analytical uncertainty is 10% for the modifier and 13% for MaxCalix, 14% for Isopar™ L, and 20% for TiDG. N/A = Not Applicable.

<sup>\*</sup> Nominal value is the expected value for freshly prepared blended solvent with a target density of 0.8352 g/mL at 25 °C.

<sup>§</sup> 
$$x = \frac{\sum_i^t \left( \frac{x_i}{\delta_i^2} \right)}{\sum_i^t \left( \frac{1}{\delta_i^2} \right)};$$
  $x_i$  stands for the concentration obtained at a given method and  $\delta_i$  is the corresponding uncertainty.



**Figure 2. Suppressor concentration as measured by titration in SHT samples since NGS implementation. The minimum recommended is 480 mg/L for TiDG.**



**Figure 3. MaxCalix concentration as measured by HPLC and FT-HNMR of recent samples since NGS implementation (44,400 mg/L is the nominal concentration).**

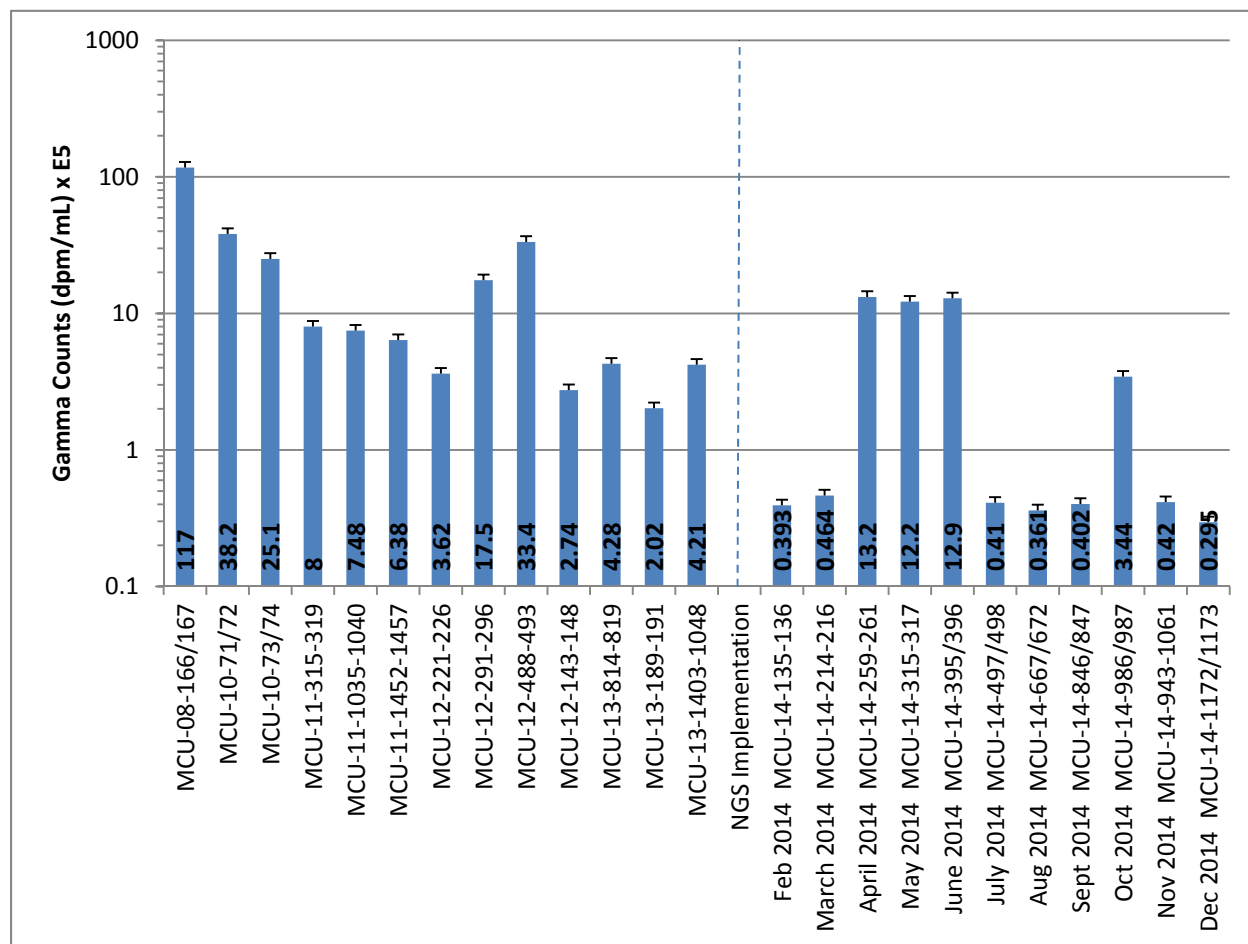


Figure 4. The gamma count of selected SHT samples. One standard deviation is 5%.

#### 4.0 Conclusions

SRNL received one set of SHT samples (MCU-14-1172-1173 pulled on 12/30/2014) for analysis. The sample was analyzed for composition. Analysis of sample MCU-14-1172-1173 indicated low concentrations of the suppressor (TiDG), of the extractant (MaxCalix), and of the modifier (CS-7SB) in the solvent relative to their nominal values. This analysis confirms a downward trend of these components some possibly due to excess Isopar™ L. No impurities were found in this solvent. The laboratory will continue to monitor the quality of the solvent in particular for any new impurity or degradation of the solvent components.

## 5.0 References

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<sup>1</sup> W. M. Matthews, HLW-CRF-10006, Rev. 0, May 18, 2010.

<sup>2</sup> T. B. Peters and M. R. Williams, “Results of Analysis of NGS Concentrate Drum Samples” SRNL-STI-2013-00521, September 2013.

<sup>3</sup> B. A Moyer, L. H. Delmau, B. D. Roach, and N. J. Williams, “Thermal Degradation of Next Generation Solvent using Triisodecylguanidine Suppressor: Impacts on Solvent Performance and Organic Content of Aqueous Effluents” ORNL-LTR-NGCSSX-020, Rev. 1, July 2013.

<sup>4</sup> K. M. L. Taylor-Pashow, F. F. Fondeur, T. L. White, D. P. Diprete, and C. E. Milliken, “Development of Analytical Methods for Determining Suppressor Concentration in the MCU Next Generation Solvent (NGS)” SRNL-STI-2013-00435, Rev. 0, July 2013.



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