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Retention: Permanent

Bioavailability of Radioactive Cesium in Old R Discharge Canal, R-Canal, Pond A, and the Adjacent Flood Plain

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September 27, 2001



Westinghouse Savannah River Company Savannah River Site Aiken, SC 29808



This document was prepared in conjunction with work accomplished under Contract No. DE-AC09-96SR18500 with the U.S. Department of Energy.

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Printed in the United States of America

Prepared For U.S. Department of Energy

Cover Photo: Cs-Illite Limnocoral Project in R-Canal. Photograph by A. Knox.

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October 27, 2001

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1.0 EXECUTIVE SUMMARY

Risk calculations are often based on the total concentration of a contaminant within the sediment, however, such calculations may over-estimate risks because not all of the contaminant is available for uptake by the biota. Determining the fraction of the total contaminant within the sediment that is not bioavailable could make risk calculations more realistic and potentially reduce the extent (source term) of required cleanup.

The objectives of this study were to provide site-specific data for human and ecological risk analysis associated with ¹³⁷Cs contamination along the P and R reactor cooling canal system. This area, which contains wetlands, was contaminated with ¹³⁷Cs during the operation of R Reactor prior to 1964. For this study, bioavailability was measured directly from ¹³⁷Cs concentrations in paired sediment and plant samples collected at the study site. These data were used to calculate concentration ratios (CR = activity concentration in the dried plant / activity concentration in dried sediment). Additionally, the nature of the bond between ¹³⁷Cs and the sediments at this site were characterized through sequential extraction techniques. These measurements provided insight into how strongly ¹³⁷Cs is bound to the sediment and provide indirect information about the tendency for ¹³⁷Cs to be taken up by plants. Among the key findings from this research are the following:

- Sequential extraction results indicated that between 50 and 85% of the ¹³⁷Cs in these sediments was strongly bound and likely in a chemical form that was of limited bioavailability.
- Estimates of concentration ratios were often in excess of 1.0, indicating that the plants concentrated ¹³⁷Cs in their tissue to levels greater than existed in the sediment.
- Concentration ratios of plants growing on terrestrial sites were significantly greater than those of the same species growing in aquatic locations.
- Mean (± SD) ¹³⁷Cs concentrations in the Old R Discharge Canal (189 ± 167 pCi g⁻¹) were no different from the average for R Canal (224 ± 213 pCi g⁻¹) or Pond A (213 ± 205 pCi g⁻¹). Part of the motivation for sampling the Old R Discharge Canal was that gamma over-flight data indicated that ¹³⁷Cs contamination levels were higher there than in Pond A. This apparent discrepancy is likely the result of increased shielding of the gamma emissions by the greater depth of water in Pond A as compared to the Old R Discharge Canal. These findings indicate that remediation decisions about ¹³⁷Cs-contaminated wetlands could be misdirected if based solely on gamma over-flight data.

Since predictions of ¹³⁷Cs concentrations in plants are tenuous when based solely on sediment ¹³⁷Cs concentrations, and the predictions are not captured well by a single parameter, such as a CR, it is recommended that risk assessments involving the ingestion pathway be based directly on measured concentrations in plants. Relying solely on sediment ¹³⁷Cs concentrations, either total or that which is biologically available, could cause large errors in risk estimations.

2.0 INTRODUCTION

2.1 BACKGROUND

Over 1,200 hectares (3,000 acres) of wetlands on the Savannah River Site (SRS) are contaminated with 21 TBq (564 Ci) of ¹³⁷Cs and pose a significant remediation challenge to the Department of Energy (DOE). The problems of contaminated aquatic systems and their associated risks are heightened on the SRS because ¹³⁷Cs is more mobile here than at any other DOE facility. Reasons for the enhanced bioavailability of ¹³⁷Cs are largely due to the low clay content of SRS sediments, and that the clays that are present are dominated by kaolinite and iron oxides. The abundance of clay is important in determining bioavailability because ¹³⁷Cs can be bound within the physical structure of clay minerals (Francis and Brinkely 1976), in essence, trapping ¹³⁷Cs and reducing its biological uptake until physical decay removes it from the system. Thus, as the percentage of clay increases, availability of ¹³⁷Cs generally decreases. Kaolinite's physical structure, however, does not bind ¹³⁷Cs as do clays that are in 2:1 layers, such as micas.

Cesium undergoes a seasonal cycle of enhanced bioavailability on the SRS. In summer, lakes on the SRS thermally stratify and the deeper hypolimnion layer becomes anoxic. During this period, ¹³⁷Cs is released. The cause for the release under anoxic conditions may be attributed to either the dissolution of Fe(III)-oxides, which sorb¹³⁷Cs, or to the microbial generation of NH₄⁺ from NO₃⁻. Ammonium has a higher affinity for the clay binding sites to which ¹³⁷Cs is adsorbed, and thus displaces ¹³⁷Cs into the interstitial waters, thereby increasing the mobility and biological availability of the contaminant. Although only a small percentage of the total inventory, the released ¹³⁷Cs is sufficient to cause measurable concentrations in all components of the wetlands (e.g. water, plants, aquatic insects, fish, etc).

Increased 137 Cs availability was documented in these systems during the 1960s era of weapons fallout (Whicker and Schultz 1982), and the high mobility of 137 Cs still exists today, as is evident by the propensity of 137 Cs to be taken up from the sediment by plants. This propensity can be measured using a concentration ratio (CR), where CR = Bq kg⁻¹ dried plant / Bq kg⁻¹ dried sediment. The National Council on Radiological Protection and Measurements recommends a default 137 Cs CR of 0.1 for use in risk calculations. In comparison, 137 Cs CRs for the SRS are generally well above 1.0. Indeed, they are among the highest recorded 137 Cs CRs in the world. Research conducted in the 1970s found plants from Steel Creek and Par Pond to have CRs that ranged from 3 to 7 (Garten and Paine 1977; Pinder et al. 1980). More recent research found that garden vegetables grown on the exposed lake bed of Par Pond had CRs of 6.8 ± 2.3 (Seels et al. 1995), and early successional species invading Par Pond's exposed lake bed had very high CRs of 14.9 ± 2.3 (Hinton et al. 1999). Such large CRs indicates how readily 137 Cs is taken up by plants and how weakly the 137 Cs is held by SRS sediments. Because of these large CRs, 137 Cs can be the critical contaminant when human and ecological risk analyses are conducted for the SRS. Part of the work reported herein uses CRs to examine the bioavailability of 137 Cs in wetlands of the SRS.

Human and ecological risk calculations will likely be used to determine the extent of remediation required within these wetlands. Often, risk calculations are based on the contaminant concentrations within the sediments. Such calculations, however, may overestimate risks because not all of the contaminant is available for uptake by the biota. Some portion of the ¹³⁷Cs is sorbed onto the sediment and is not readily taken up by plants or dissolved into the overlying waters. Determining the available fraction of the total contaminant within the sediment (i.e. that which is available for plant uptake) could make risk calculations more realistic and potentially reduce the extent of required cleanup. More directly, the bioavailable fraction could be based directly on the amount of contaminant measured in field samples. Remediation of sensitive wetlands is expensive and is also an extremely difficult procedure to accomplish without destroying the fragile ecosystems.

Use of total concentration as a criterion to assess the potential effects of Cs in the sediment environment is generally not satisfactory. To provide a more precise understanding of the potential impact of elevated levels of Cs in sediments, it is necessary to identify and quantify the forms in which Cs is present in the sediment. Sequential extractions are often used to estimate the available fraction of a contaminant. Sequential extractions are used to examine the solubility of a contaminant in progressively stronger reagents. The reagents are chosen to target specific components within the sediment matrix that the contaminant may be residing in. Part of the work reported herein uses sequential extractions to examine the distribution of ¹³⁷Cs in SRS wetland sediments.

This study was conducted at R-Canal, Pond A, and the Old R Discharge Canal because they have been shown to contain contaminant ¹³⁷Cs. This area consists of wetlands, ponds, and canals with standing water that varies from a few centimeters to 150 cm in depth. The wetlands' associated flood plains were also studied and are referred to in this report as terrestrial sites.

2.2 OBJECTIVES AND SCOPE

This research was designed to improve site-specific risk calculations from ¹³⁷Cs-contaminated wetlands on the SRS. Risk calculations based on total ¹³⁷Cs concentrations in sediments may lead to an over-estimation of hazards, unnecessary destruction of sensitive wetlands, and needless expenditure of funds.

The objective of this proposal was to provide more realistic data for human and ecological risk analysis. Specifically, the objectives and scope of this study were to:

- collect plant and associated sediment samples from terrestrial flood plain sites on the Old R Discharge Canal and determine bioavailability of ¹³⁷Cs to specific plant species based on concentration ratios,
- collect plant and associated sediment samples from aquatic sites of the Old R
 Discharge Canal and to compare ¹³⁷Cs bioavailability in the terrestrial flood plain and
 aquatic systems,
- determine, through the use of sequential extraction techniques, which specific mineral phases bind ¹³⁷Cs, and

• estimate availability of ¹³⁷Cs to aquatic and terrestrial plants using concentration ratios.

3.0 MATERIALS AND METHODS

3.1 SEDIMENT CHARACTERIZATION

A total of 12 sampling locations were randomly selected along the Old R Discharge Canal located ~ 0.8 km along the cooling water outflow canal from R- reactor (Figure 1). At each location, both plant and associated sediment samples from aquatic and terrestrial flood plain sites were collected. In June, sixteen plant and sediment samples were taken from the terrestrial sites and 14 from the aquatic (Figure 1 and Table 1). Sediment samples were collected from the rooting zone of the plants. Most sediment samples were collected as a grab sample (0-7.5 cm in depth), but sediment cores were taken from 5 aquatic sites. The sediment cores were sliced into 2.5 cm sections and each slice was processed as an individual sample. Sediment samples were oven dried (60-65°C), homogenized, and sieved through a 2-mm screen. Sediment pH was determined from a 1:1 sediment/water equilibration solution (Thomas, 1996). Percent soil organic matter was estimated by loss-on–ignition at a temperature of 450 °C. Particle size distributions (percent sand, silt, and clay) were determined by the micro-pipette method (Miller and Miller, 1987). Sediment samples with organic matter content higher than 10% had their particle size distribution analysis performed after organic matter was removed with H₂O₂.

As part of another study on R-Canal and Pond A, 16 limnocorrals have been established to study the effects of illite clay amendments on Cs mobility and bioavailability (Figure 2). Limnocorrals are flexible, plastic cylinders (3 m in diameter) that are anchored to the pond sediment and extend upwards through the water column to the atmosphere. We used these limnocorrals as additional sediment/plant sampling locations for the study reported herein. From each limnocorral three replicate sediment cores were taken.

3.2 PLANT SAMPLES

Plant species taken from the Old R Discharge Canal were royal fern (*Osmunda regalis*), netted chain fern (*Woodwardia aerloata*), and rice cutgrass (*Leersia oryzoides*) (Table 1). These species were selected because they are abundant in the area and because they were found in both aquatic and terrestrial environments. From the limnocorrals in Pond A, we collected three replicate samples of water lily (*Nymphea odorata*) leaves and stems. All plant samples were cleaned with tap water and oven-dried at 60-65°C. Plant samples were then ground, weighed, and placed into 24-ml scintillation vials for ¹³⁷Cs analyses.

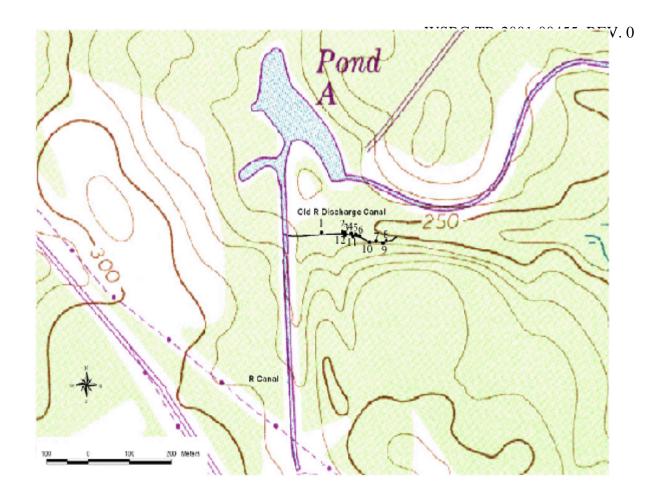


Figure 1. Old R Discharge Canal's position relative to Pond A and the main R-Canal. Dots indicate 12 sampling locations. Contour lines depict topographical elevation.

3.3 ¹³⁷Cs ANALYSES

Analyses of ¹³⁷Cs in plants, sediments, and sequential extracts were performed on a Packard Autogamma instrument with a well-type NaI crystal (7.6 x 8.3 cm with a 1.5 % relative efficiency). Additional analyses of portions of the sequential extracts were performed on a 70x90 mm² high-purity germanium detector. Cs-137 standards, traceable to the National Institute of Standards and Technology, were assayed in the same physical geometry as the samples.

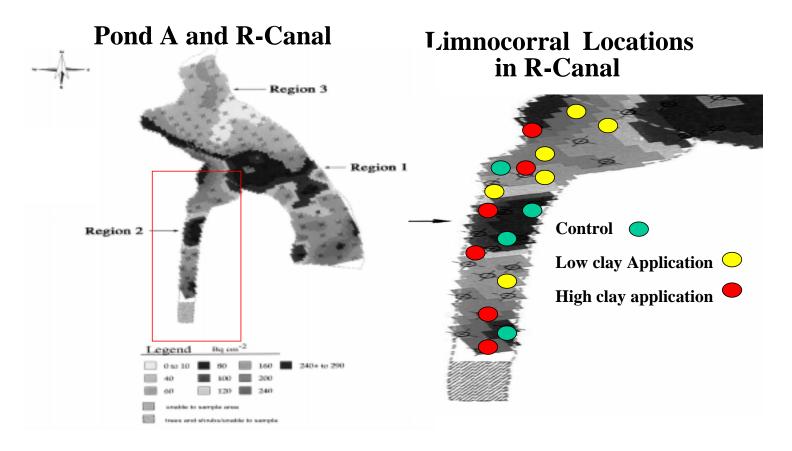


Figure 2. Location of 16 limnocorrals in R-Canal from which sediment and water lily samples were taken.

Table 1. Plant samples collected at Old R Discharge Canal; T- terrestrial, A- aquatic.

-	Roya	al Fern	Netted (Chain Fern	Rice (Cutgrass
Location	Aquatic	Terrestrial	Aquatic	Terrestrial	Aquatic	Terrestrial
1	-	X	-	-	-	-
2	X	_	_	-	_	-
	-	${f X}$	-	-		
3	-	-	-	-	\mathbf{X}	-
	-	-	-	-	-	\mathbf{X}
4	X	-	-	-	-	-
	-	-	-	-	\mathbf{X}	-
	-	-	-	-	-	\mathbf{X}
5	\mathbf{X}	-	-	-	-	-
	_	_	_	-	\mathbf{X}	-
	_	${f X}$	_	-	_	-
	-	-	-	-	-	\mathbf{X}
6	-	-	-	${f X}$	-	-
	-	-	-	-	-	${f X}$
7	-	-	-	-	\mathbf{X}	-
	-	-	-	${f X}$	-	-
8	-	-	\mathbf{X}	-	-	-
	-	-	-	-	\mathbf{X}	-
	-	-	-	${f X}$	-	-
	-	-	-	-	-	\mathbf{X}
9	\mathbf{X}	-	-	-	-	-
	-	-	-	-	\mathbf{X}	-
	-	-	-	${f X}$	-	-
	-	-	-	\mathbf{X}	-	-
10	X	-	-	-	-	-
	-	-	-	-	-	\mathbf{X}
11	-	-	-	-	\mathbf{X}	-
	-	-	-	${f X}$	-	-
12	-	-	\mathbf{X}	-	-	-
	-	-	-	X	-	
Number of Samples	5	3	2	7	7	6
Total Number of		_		_		
Samples/Species		8		9	-	13

3.4 SEQUENTIAL EXTRA CTIONS

Chemical extraction of sediment contaminants with sequentially more aggressive reagents is commonly used to characterize various forms of metal or radionuclides in sediments. Numerous sequential extraction procedures have been developed (Tessier et al., 1979, Miller et al., 1986; Hall et al., 1995). The major problem with sequential extraction studies is that the detailed nature of the material being extracted is not known. Idealized pure phases are often named in various schemes on the basis of their response to the extraction reagents, but this approach can be misleading in that the behavior of natural samples may be variable depending on the inorganic and organic constituents with which metals are associated. Spectroscopic methods may yield more precise information regarding the identification of the metal forms, the specific phases the metal is bound to, and the nature of the sedimentmetal bond (e.g., bond strength, bond distance, and bond type). But these methods are usually expensive, non-routine, and not sensitive enough to detect contaminants in low concentrations. Therefore, sequential extractions, even though they have several limitations, are still widely used.

A sequential extraction procedure, modified from Hall et al. (1995) and Tessier et al. (1979), was conducted on eight sediments collected from R-Canal and Pond A. An overview of the sequential extraction procedure used in this study is presented in Table 2. The technique differentiates five sediment fractions in which ¹³⁷Cs was found:

- Adsorbed/Exchangeable/Carbonate (AEC),
- Amorphous Fe-oxyhydroxide (AmFeOx),
- Crystalline Fe-oxide (CryFeOx),
- Organic Matter (OM), and
- Residual silicates (R).

Exchangeable Cs is held by electrostatic attraction on the surface of negatively charged sediment complexes. Neutral salts such as MgCl2 or ammonium acetate can easily displace cesium held to exchange sites. This fraction is often considered the most biologically available. In the sequential extraction methods we used, exchangeable and carbonate bound (not abundant in SRS sediments) Cs was grouped together. This fraction was extracted with sodium acetate, buffered at pH 5, and designated the Adsorbed/Exchangeable/Carbonate complex (AEC). Also extracted together, in the next step, were the hydrous oxides of Mn and Fe, well known "sinks" in the surficial environment for heavy metals and radionuclides. Scavenging by these secondary oxides, present as coatings on mineral surface or as fine discrete particles, may occur by any combination of the following mechanisms: coprecipitation, adsorption, surface complex formation, ion exchange, and/or penetration of the mineral lattice. The third fraction consisted of crystalline Fe oxides. The relative importance of Fe and Mn oxides as scavengers depends upon such conditions as pH-Eh values, degree of oxide crystallization, their abundances, and the presence of organic matter. Cesium bound to organic matter was next in the sequence and represented the fourth extract. The final part of the sequential extraction procedure represented the residual fraction, and is

thought to mainly consist of primary and secondary silicate minerals that may hold Cs tenaciously within their crystal structure. This fraction was determined by gamma spectroscopy after the four prior extraction steps.

Sediment cores from R-Canal and Pond A were sliced into 2.5 cm sections and each slice was characterized for total ¹³⁷Cs, pH, and particle size distribution as described in Section 3.1. Eight representative sediment samples from R-Canal were selected for sequential extraction (two samples with high organic matter content, two samples with high clay content, two samples with high content of organic matter and clay, and three samples with high content of sand fraction). All extracts were performed by shaking the samples in 50 ml centrifuge tubes with the appropriate reagents at a 40:2 reagent to sample ratio, and spun for 30 min at 2800 rpm. The extracts were transferred to two 20 ml vials in preparation for determining ¹³⁷Cs concentration. The residue was prepared for the next extraction step by washing with 10 ml of 0.01 M CaCl₂ (three times), centrifuging at 2800 rpm for 30 min and collecting the decanted supernatant liquid into a vial for ¹³⁷Cs analysis. ¹³⁷Cs activity was measured directly in the remaining solid phase (as opposed to after total digestion by strong acids, such as aqua regia).

Table 2. Sequential extraction regime used in this study.

Sequence	Extractant: Chemical Composition	Targeted Contaminant Fraction	Ref. ^a
Initial sample	2 g of sample	Total ¹³⁷ Cs activity of the sample was performed on high-purity geranium detector	
1	1.0 M NaOAc; pH 5; repeated twice	d Adsorbed/exchangeable/carbonate (AEC)	1
2	0.25 M NH ₂ OH·HCl in 0.25 M HCl; repeated twice	Amorphous Fe oxyhydroxide (Am Fe ox)	1
3	1.0 M NH ₂ OH·HCl in 25% HOAc; repeated twice	Crystalline Fe oxide (Cry Fe ox)	1
4	0.02 N HNO ₃ & 30% H ₂ O ₂ ; repeated twice	Organic Fraction (OM)	2
5		Residual (silicates) (R)	

^a References: 1 = Hall et al. 1995, 2 = Tessier et al. 1979.

4.0 RESULTS AND DISCUSSION

4.1 SEDIMENT CHARACTERIZATION

Sediments collected at aquatic sites had a pH range of 3.9 to 5.1 with a mean of 4.3 (Table 3). Organic matter content was high in five samples (more than 5% but less than 10%, wt) and very high in seven samples (more than 20%, wt; Table 3). A high content of organic matter is consistent with the fact that the sediments were collected from a forested wetland. Particle size analysis showed that the textures of the sediments from aquatic sites varied substantially (Table 3). The clay-size fraction was the smallest and ranged from 0.5 to 8.9%, with a mean of 3.8%. The silt-size fraction varied substantially, ranging from 0.7 to 85%, with a mean of 44.1%. The highest fraction for aquatic sediments was sand with a mean of 52% (Table 3).

Sediment samples collected from the terrestrial flood plain sites were remarkably similar to those collected from the aquatic sites in sand and silt content (p > 0.05); but the clay content was significantly lower in the terrestrial site (p < 0.01), with a mean percentage of 1.5 (Table 4). The two types of sites had similar pH values and organic matter content (Table 4).

Table 3. Sediment characterization of samples collected from aquatic locations in the Old R Discharge Canal.

		quatic sediment sa	mples			
Location	Associated plant sample	OM	pН	Sand	Silt	Clay
		(%, wt)	(unitless)	(%, wt)	(%, wt)	(%, wt)
2	Royal Fern	6.65	3.95	68	29	3
3	Rice Cutgrass	6.41	4.25	52	44	5
4	Royal Fern	7.86	4.35	76	22	2
4	Rice Cutgrass	7.17	4.22	50	45	5
5	Rice Cutgrass	29.11	5.1	29	67	5
5	Royal Fern	12.33	5	6	85	9
7	Rice Cutgrass	22.40	4.22	7	85	8
8	Netted Chain Fern	61.25	4.17	98	1	1
8	Rice Cutgrass	33.60	3.85	63	34	2
9	Royal Fern	7.00	4.23	64	32	3
9	Rice Cutgrass	6.91	4.18	84	15	1
10	Royal Fern	43.42	4.17			
11	Rice Cutgrass	47.14	3.88	53	45	2
12	Netted Chain Fern	29.03	4.54	28	70	3
	Mean	22.88	4.42	52	44	4
	Standard Deviation	± 18.19	± 0.45	± 28	± 26	± 3

Table 4. Sediment characterization of samples collected from terrestrial flood plain sites along the Old R Discharge Canal.

	161	restrial sediment s OM	pH	Sand	Silt	Clay
Location	Associated plant sample	(%, wt)	(unitless)	(%, wt)	(%, wt)	(%, wt)
1	Royal Fern	14.04	3.61	45	50	5
2	Royal Fern	53.00	3.89	63	36	1
3	Rice Cutgrass	7.07	5.43	80	20	0
4	Rice Cutgrass	20.84	3.92	78	22	1
5	Royal fern	33.07	4.72	85	22	0
5	Rice Cutgrass	29.03	5.35	55	45	1
6	Rice Cutgrass	16.66	4.3	90	10	0
6	Netted Chain Fern	69.49	3.61	36	63	1
7	Netted Chain Fern	50.33	4.09	30	68	3
8	Netted Chain Fern	19.53	4.06	72	26	2
8	Rice Cutgrass	10.12	4.31	75	23	2
9	Netted Chain Fern	14.86	3.76	49	50	4
9	Netted Chain Fern	19.50	3.98	82	17	2
10	Rice Cutgrass	3.44	5.65	83	16	0
11	Netted Chain Fern	14.07	5.06	87	13	0
12	Netted Chain Fern	7.96	4.64			
Av	erage	23.94	4.40	67	32	2
Std	ev	18.72	0.67	20	19	2

4.2 SEDIMENT ¹³⁷Cs CONCENTRATIONS

¹³⁷Cs concentrations in the sediments from the terrestrial and the aquatic sites of Old R Discharge Canal are presented in Table 5. The highest reported ¹³⁷Cs concentrations were in the sediment samples collected at the aquatic sites, e.g., 13.8 and 24.3 Bq g⁻¹ (373 and 656 pCi g⁻¹). The mean (\pm SD) concentration of ¹³⁷Cs in the sediments from the aquatic sites was 7.0 \pm 6.2 Bq g⁻¹ (189 \pm 167 pCi g⁻¹). In contrast, ¹³⁷Cs concentrations from the terrestrial flood plain sites were considerably lower, with a mean (\pm SD) of 1.3 \pm 1.8 Bq g⁻¹ (35 \pm 49 pCi g⁻¹) and a range from 0.08 to 6.2 Bq g⁻¹ (2 to 167 pCi g⁻¹).

 137 Cs concentrations in the sediment cores taken from R-Canal and Pond A are reported in Table 6. The concentrations of 137 Cs in the sediment samples change with depth. On average, the greatest 137 Cs concentrations (9.0 \pm 6.9 Bq g $^{-1}$; 243 \pm 186 pCi g $^{-1}$) were measured 2.5 to 5.0 cm deep within the sediments, and concentrations then decreased with depth. This pattern is consistent with earlier published data from Pond A (Abraham et al., 2000).

The mean $(\pm \, SD)^{137}$ Cs concentration in the Old R Discharge Canal $(7.0 \pm 6.2 \, Bq \, g^{-1}; \, 189 \pm 167 \, pCi \, g^{-1})$ was no different from the mean for R Canal $(8.3 \pm 7.9 \, Bq \, g^{-1}; \, 224 \pm 213 \, pCi \, g^{-1})$ or from the mean reported for Pond A $(7.9 \pm 7.6 \, Bq \, g^{-1}; \, 213 \pm 205 \, pCi \, g^{-1})$ by Abraham et al. (2000) (Figure 3). Part of the motivation for sampling the Old R Discharge Canal was that gamma over-flight data indicated that 137 Cs contamination levels were higher there than in Pond A. This apparent discrepancy is likely the result of increased shielding of the gamma emissions by the greater depth of water in Pond A as compared to the Old R Discharge Canal. These findings indicate that remediation decisions about 137 Cs-contaminated wetlands could be misdirected if based solely on gamma over-flight data.

4.3 SEQUENTIAL EXTRA CTIONS

The sediment samples studied by sequential extractions represent four "typical sediment types" found in Pond A and the R-Canal system; sediments with:

- 1) high organic matter content,
- 2) high clay content,
- 3) a combination of high clay and organic matter content, and
- 4) high sand content.

All 48 cores (3 replicates x 16 locations) collected from the limnocorral study conducted in R-Canal contained various proportions of these "typical sediment types." These sediment classifications were easy to identify by unaided visual inspection of the cores. The sequential extraction technique allowed us to determine ¹³⁷Cs concentrations in five components of each sediment. The components (AEC, AmFeOx, CryFeOx, OM and R) were presented and discussed in section 3.4. ¹³⁷Cs was found in each component of all sediment types analyzed

(Figure 4). In all four "typical sediment types," the greatest concentration of ¹³⁷Cs was found in the AmFeOx component (Figure 4). When compared across sediment types, the AEC component held a larger percentage (24%) of the ¹³⁷Cs in the sand than in the other three sediment types (Figure 4). Recall that the AEC component is the mineral matrix where Cs is most easily exchanged with other elements, and thus considered by some to be an estimate of the biologically available fraction. This fraction was greater than 15% in all sediment types. The residual fraction is expected to bind and hold Cs tenaciously, thereby reducing its bioavailability. As expected, we found the clay sediment type to contain the most ¹³⁷Cs in the residual fraction and sand to have the least (Figure 4).

Overall, the sequential extractions results suggest that anywhere from 10 to 50% of the total ¹³⁷Cs within the sediments of Pond A might be considered biologically available—depending on the seasonal site specific conditions within the sediments.

4.4 PLANT ¹³⁷Cs CONCENTRATIONS

Within the Old R Discharge Canal three plant species were collected: royal fern (Osmunda regalis), netted chain fern (Woodwardia aerloata), and rice cutgrass (Leersia oryzoides). The same species were collected at the aquatic and terrestrial flood plain sites (Table 5). The highest ¹³⁷Cs concentrations were observed in netted chain fern at an aquatic site; 8.1 Bq g⁻¹ (219 pCi g⁻¹). Because ¹³⁷Cs concentrations in sediments from aquatic sites were significantly higher, 7.0 ± 6.1 Bq g⁻¹ (189 ± 165 pCi g⁻¹), than those from the terrestrial flood plain, 1.3 ± 1.8 Bq g⁻¹ (35 ± 49 pCi g⁻¹) one might expect ¹³⁷Cs concentrations in plants to be higher in aquatic sites as well. However, the mean ¹³⁷Cs concentrations in all plants collected from aquatic and terrestrial flood plain sites were similar, 1.9 ± 2.1 and 1.7 ± 2.1 Bq g^{-1} , $(51 \pm 57 \text{ and } 46 \pm 57 \text{ pCi g}^{-1})$. Some researchers have shown that sediment ¹³⁷Cs activity concentrations were positively correlated to plant uptake (Pinder et al., 1980), while others were unable to identify such a relationship (Horrill et al;, 1990), indicating that plant concentrations do not necessarily depend upon sediment concentrations. Hughes et al., (1980) indicated that the main factors influencing plant uptake are sediment pH, and level and chemical species of contaminants. The influence of plant species on metal uptake may also be considerable. Different species regulate metal uptake at both the sediment-root and root-shoot interfaces to varying degrees (Archer, 1971). This was also observed in our data, in that ¹³⁷Cs concentrations in rice cutgrass were consistently lower than the two fern species (Table 5). Ferns are known for their tendency to take up radionuclides and heavy metals (Kaplan et al. 2001).

At R-Canal and Pond A, water lilies were collected at 16 locations. Generally, ¹³⁷Cs concentrations were higher in stems than in leaves. In leaves ¹³⁷Cs concentrations ranged from 0.9 to 1.5 Bq g⁻¹ (24 to 40 pCi g⁻¹), while in stems ¹³⁷Cs concentrations were almost two times higher and ranged from 1.2 to 2.8 Bq g⁻¹ (32 to 76 pCi g⁻¹, Table 7).

Table 5. ¹³⁷Cs concentrations in sediments, concentrations in associated plants, and concentration ratios (CR) from aquatic and terrestrial sites of Old R Discharge Canal.

Aquatic Site					Terrestrial Site					
		¹³⁷ Cs in	¹³⁷ Cs in				¹³⁷ Cs in			
		sediments	plant	CR			sediments	¹³⁷ Cs in plant	CR	
Location	Sampled plant	(Bq g ⁻¹)	(Bq g ⁻¹)		Location	Sampled plant	(Bq g ⁻¹)	(Bq g ⁻¹)		
2	Royal Fern	7.40	4.55	0.61	1	Royal Fern	3.52	5.90	1.68	
3	Rice Cutgrass	9.23	0.87	0.09	2	Royal Fern	1.88	2.72	1.45	
4	Royal Fern	4.64	0.86	0.18	3	Rice Cutgrass	0.16	0.03	0.17	
4	Rice Cutgrass	10.09	1.00	0.09	4	Rice Cutgrass	0.13	0.02	0.11	
5	Rice Cutgrass	3.28	0.78	0.23	5	Royal fern	3.62	2.16	0.60	
5	Royal Fern	13.82	0.61	0.04	5	Rice Cutgrass	6.22	0.60	0.10	
7	Rice Cutgrass	24.33	1.29	0.05	6	Rice Cutgrass	0.09	0.06	0.61	
8	Netted Chain Fern	7.44	8.05	1.08	6 Netted Chain Fern		1.47	4.77	3.24	
8	Rice Cutgrass	2.74	1.34	0.49	7	Netted Chain Fern	0.82	0.93	1.13	
9	Royal Fern	2.17	1.57	0.72	8	Netted Chain Fern	0.18	1.27	6.88	
9	Rice Cutgrass	0.13	0.20	1.59	8	Rice Cutgrass	0.14	1.35	9.45	
10	Royal Fern	5.14	2.87	0.56	9	Netted Chain Fern	0.17	0.52	3.07	
11	Rice Cutgrass	3.98	1.45	0.36	9	Netted Chain Fern	0.05	0.33	6.57	
12	Netted Chain Fern	3.66	0.78	0.21	10	Rice Cutgrass	0.08	0.03	0.44	
					11	Netted Chain Fern	2.28	5.78	2.54	
					12	Netted Chain Fern	0.08	0.24	2.93	
Overall Mean	1	7.00	1.88	0.45	Overall Me	ean	1.31	1.67	2.56	
Overall Stand	lard Deviation	6.16	2.09	0.45	Overall Sta	ndard Deviation	1.80	2.06	2.79	
Mean (±SD) Royal Fern	6.6 ± 4.4	2.1 ± 1.6	0.4 ± 0.3	Mean (±S	D) Royal Fern	3.0 ± 0.9	3.6 ± 2.0	1.2 ± 0.6	
Mean (±SD	Netted Chain Fern	5.5 ± 2.6	4.4 ± 5.1	0.6 ± 0.6	Mean (±S	D) Netted Chain Fern	1.0 ± 0.9	2.0 ± 2.3	3.8 ± 2.2	
Mean (±SD) Rice Cutgrass	8.5 ± 8.6	1.0 ± 4.3	0.4 ± 0.5	Mean (±S	D) Rice Cutgrass	1.1 ± 2.5	0.4 ± 0.6	1.9 ± 2.6	

Table 6. Concentration of 137 Cs (Bq g $^{-1}$) as a function of depth in sediment cores collected at R-Canal and Pond A. Blanks within the table mean that the core was shallow and not taken to the depth indicated. Values can be multiplied by 27 to obtain concentrations in units of pCi g $^{-1}$.

		Dept	h of Core Slice	(cm)	
Site	0-2.5	2.5-5.0	5.0-7.5	7.5-10	10-12.5
1	7.3	8.06	8.3	12.3	10.2
2	16.9	6.4	6.1	-	-
3	2.2	5.9	4.2	3.4	2.4
4	24.1	19.2	10.9	-	-
5	2.0	6.3	-	-	-
6	7.6	19.0	45.7	42.5	29.1
7	2.0	1.7	2.3	1.8	-
8	2.0	19.2	1.9	1.4	1.2
9	1.6	2.8	8.1	36.5	31.4
10	2.4	14.8	8.0	2.8	1.0
11	2.3	16.0	3.3	-	-
12	0.7	1.1	1.9	0.9	-
13	3.0	3.6	9.5	2.5	0.6
14	1.3	1.9	1.5	1.7	1.8
15	1.1	1.4	1.9	12.4	12.9
16	1.9	21.0	9.7	3.8	-
17	1.9	4.3	8.0	5.0	2.8
18	2.8	13.3	4.6	1.4	1.2
19	5.1	9.6	16.6	10.0	6.6
20	1.7	2.2	1.9	-	-
21	1.9	5.5	1.7	1.5	1.1
22	59.6	6.1	4.6	2.0	-
23	3.9	1.7	0.3	0.1	-
24	1.8	4.7	-	-	-
25	3.6	19.7	7.9	2.8	3.5
26	9.2	19.2	4.6	1.7	0.6
27	1.6	13.3	2.5	0.7	0.5
28	3.5	11.8	4.3	2.6	1.9
29	1.8	1.7	1.8	3.7	6.3
Mean	6.1	9.0	6.7	6.7	6.4
SD	11.5	6.9	8.6	10.9	9.4

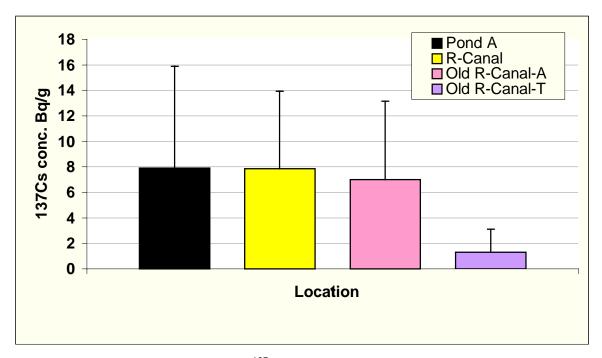


Figure 3. Average concentrations of ¹³⁷Cs (Bq g-1) in sediment at Pond A, R-Canal and Old R-Canal. Data for Pond A are from Abraham et al. (2000). Data for both aquatic (A) and terrestrial sites (T) are reported for Old R-Canal. Values can be multiplied by 27 to obtain results in units of pCi g⁻¹.

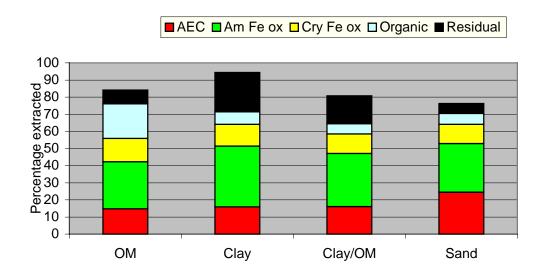


Figure 4. Partition of ¹³⁷Cs among five fractions in four types of sediment from R-Canal and Pond A; AEC – adsorbed/exchangeable/carbonate phase, Am Fe ox – amorphous Fe oxyhydroxide, Cry Fe ox – crystalline Fe oxide, Organic – organic phase, Residual – silicates.

 $\textbf{Table 7.} \ ^{137}\!Cs \ concentrations \ in \ leaves \ and \ stems \ of \ water \ lily \ collected \ at \ R-Canal \ and \ Pond \ A.$

Leaf ¹³⁷ Cs Concentrations (Bq g ⁻¹)					Stem ¹³⁷ Cs Concentrations (Bq g ⁻¹)						
Location	Rep 1	Rep 2	Rep 3	Average	Stdev.	Location	Rep 1	Rep 2	Rep 3	Average	Stdev.
1	1.53	1.18	1.69	1.46	0.26	1	2.20	1.90	2.95	2.35	0.54
2	1.54	1.71	1.28	1.51	0.21	2	2.62	3.19	2.44	2.75	0.39
3	1.14	1.47	1.45	1.35	0.18	3	1.74	1.50	1.87	1.70	0.18
4	1.09	1.05	1.55	1.23	0.27	4	1.82	2.07	2.24	2.04	0.21
5	1.18	1.39	1.02	1.20	0.18	5	1.96	2.16	2.23	2.12	0.13
6	0.80	1.30	0.94	1.01	0.25	6	1.20	1.58	1.37	1.39	0.19
7	1.10	1.28	0.95	1.11	0.16	7	1.98	4.24	1.63	2.62	1.41
8	1.0	1.09		1.07	0.02	8	2.30	1.81		2.05	0.35
9	1.29	1.11	1.20	1.20	0.09	9	2.77	1.85	2.31	2.31	0.46
10	0.87	1.05	0.89	0.94	0.10	10	0.58	1.76	1.26	1.20	0.59
11	0.97	1.07	1.11	1.05	0.07	11	1.72	1.81	2.45	1.99	0.40
12	1.28	1.33	1.10	1.24	0.11	12	1.92	2.35	1.69	1.99	0.33
13	1.02	0.98	0.92	0.97	0.04	13	1.25	1.41	1.52	1.39	0.13
14	0.77	0.90	0.89	0.85	0.07	14	1.55	1.66	1.91	1.71	0.18
15	1.22	0.81	0.89	0.97	0.21	15	1.32	1.48	1.55	1.45	0.11
16	0.91	0.89	0.73	0.85	0.09	16	1.69	1.28	1.48	1.48	0.20
Mean				1.13	0.24					1.91	0.60

4.5 137 Cs BIOAVAILABILITY – CONCENTRATION RATIOS

In addition to determining activity concentrations of ¹³⁷Cs in the plants (Bq g⁻¹ dry plant), ¹³⁷Cs uptake was normalized for the varying sediment levels by calculating concentrations ratios (CR = activity in dried plant / activity in dried sediment). The concentration ratios were calculated separately for each paired plant/sediment sample, and then the ratios were averaged across plants within each species. It is important to normalize the plant contaminant concentrations in this manner to provide insight as to the cause for low plant contaminant concentrations. Low plant contaminant concentrations could be the result of low sediment contaminant concentrations, the contaminant existing in a non-biologically available form, or avoidance of the sediment contaminant by plant roots. By expressing the plant uptake as concentrations ratios, it is possible to rule out the first cause, that is, that sediment contaminant concentrations were low.

The greatest average concentration ratio was obtained for netted fern from the terrestrial flood plain (3.8, Table 5). The average concentration ratios for water lily from R-Canal and Pond A for leaves and stems were 0.18 and 0.3, respectively (Fig. 5: Table 8). At the Old R Discharge Canal, the average 137 Cs concentration ratios for the three plant species recovered from the aquatic sites were 0.4 ± 0.4 and for the terrestrial flood plain sites were 2.6 ± 2.8 . This indicates that the CRs from plants collected in terrestrial sites were higher (Table 5). Higher concentration ratios for plants growing in terrestrial sites do not follow the findings of previous research by Hinton et al. (1999). They found CR values to be lower for terrestrial species than for wetland species (plants collected at Par Pond, SRS, Aiken SC), 8.1 and 14.8, respectively. Concentration ratios from Hinton's study are among the largest reported in the literature. Some recorded values were as high as 365, and the overall arithmetic mean for all species across 3 years was 12.5 (n = 239). Hinton et al. (1999) were studying Cs uptake in a contaminated reservoir whose water levels had recently decreased by 50%. They attributed the high CRs they observed, in part, to the conditions during the draw down of the reservoir. The species studied differed from those reported here.

Even though the CR values in this study were not as high as what Hinton et al. (1999) reported, they were 4.5 and 25.6 times greater (for aquatic and terrestrial sites) than what the National Council on Radiological Protection and Measurements recommend for a default value (0.1; NCRP, 1989). This points out, yet again, that Cs mobility is greater on the SRS than at most other sites.

Table 8. 137 Cs concentration ratios (Bq kg $^{-1}$ dried plant / Bq kg $^{-1}$ dried sediment) for water lily samples collected at R-canal and Pond A. Sediment concentrations of 137 Cs were averaged from the 0 - 12.5 cm depth.

Location	¹³⁷ Cs Concentration Ratios of Leaves						¹³⁷ Cs Concentration Ratios of Stems				
	Rep 1	Rep 2	Rep 3	Average	Stdev	Rep 1	Rep 2	Rep 3	Average	Stdev	
1	0.058	0.045	0.0641	0.056	0.010	0.084	0.072	0.112	0.089	0.021	
2	0.134	0.148	0.1115	0.131	0.019	0.227	0.277	0.212	0.239	0.034	
3	0.123	0.158	0.1564	0.146	0.020	0.187	0.162	0.202	0.184	0.020	
4	0.102	0.098	0.1439	0.114	0.026	0.169	0.192	0.208	0.190	0.019	
5	0.223	0.263	0.1940	0.227	0.035	0.371	0.410	0.421	0.401	0.026	
6	0.080	0.130	0.0944	0.102	0.026	0.121	0.159	0.138	0.139	0.019	
7	0.371	0.432	0.3223	0.375	0.055	0.671	1.432	0.550	0.884	0.478	
8	0.289	0.297	0.0000	0.196	0.169	0.628	0.493	0.000	0.374	0.331	
9	0.092	0.079	0.0849	0.085	0.007	0.196	0.131	0.164	0.164	0.033	
10	0.087	0.105	0.0891	0.094	0.010	0.058	0.176	0.126	0.120	0.059	
11	0.171	0.189	0.1951	0.185	0.013	0.302	0.318	0.431	0.350	0.070	
12	0.178	0.185	0.1535	0.172	0.016	0.266	0.327	0.235	0.276	0.047	
13	0.217	0.208	0.1964	0.207	0.010	0.265	0.299	0.323	0.296	0.029	
14	0.201	0.235	0.2311	0.222	0.019	0.403	0.433	0.497	0.444	0.048	
15	0.370	0.248	0.2718	0.297	0.065	0.400	0.449	0.470	0.440	0.036	
16	0.208	0.204	0.1680	0.193	0.022	0.386	0.292	0.337	0.338	0.047	
Mean	0.181	0.189	0.155	0.175	0.033	0.296	0.351	0.277	0.308	0.082	
SD	0.097	0.096	0.082	0.083	0.040	0.176	0.314	0.160	0.192	0.129	

5.0 SUMMARY

The objectives of this study were to provide more site-specific data for human and ecological risk analysis on the SRS. In this study the mobile fraction of ¹³⁷Cs, i.e., the fraction that could be taken up by plants, in sediments collected from R-Canal and Pond A was determined by sequential extractions of the sediments, and by directly measuring plant uptake. The propensity for plants to take up ¹³⁷Cs was calculated using a concentration ratio (CR = activity per dried plant / activity per dried sediment).

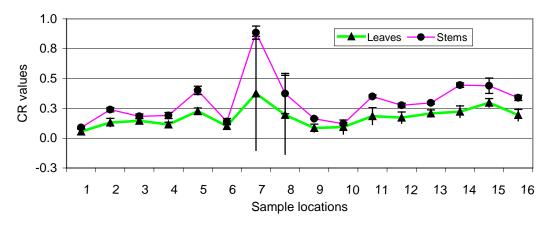


Figure 5. Mean $(\pm SD)^{137}$ Cs concentration ratios for water lily leaves and stems collected at R-Canal and Pond A; 3 replicates per location.

The study was conducted in R Canal, Pond A, and the Old R Discharge Canal. All these systems are considered wetlands and were contaminated with ¹³⁷Cs when R reactor was operating (prior to 1964).

The mean $(\pm \text{SD})^{137}\text{Cs}$ concentration in the Old R Discharge Canal $(189 \pm 167 \text{ pCi g}^{-1})$ was no different from the mean for R Canal $(224 \pm 213 \text{ pCi g}^{-1})$ or Pond A $(213 \pm 205 \text{ pCi g}^{-1})$. Part of the motivation for sampling the Old R Discharge Canal was that gamma over-flight data indicated that ^{137}Cs contamination levels were higher there than in Pond A. This apparent discrepancy is likely the result of increased shielding of the gamma emissions by the greater depth of water in Pond A as compared to the Old R Discharge Canal. These findings indicate that remediation decisions about ^{137}Cs -contaminated wetlands could be misdirected if based solely on gamma over flight data.

Sequential extraction data indicated that between 50 and 85% of the ¹³⁷Cs in these sediments was strongly bond and likely in a chemical form that was of limited bioavailability.

Estimates of concentration ratios were often in excess of 1.0, indicating that the plants concentrated ¹³⁷Cs in their tissue to levels greater than existed in the sediment. A review of

the literature indicated that ¹³⁷Cs concentration ratios >1.0 are common at the SRS, and have been attributed, in part, to the low abundance of Cs-binding clays.

Since predictions of plant ¹³⁷Cs concentrations based on sediment ¹³⁷Cs concentrations is tenuous and not captured well by a single parameter, such as a concentration ratio, it is recommended that risk assessment related to plants be based directly on plant ¹³⁷Cs concentrations. Relying solely on sediment ¹³⁷Cs concentration data, either total or that which is biologically available, could result in large errors in risk estimations.

6.0 ACKNOWLEDGMENTS

We would like to acknowledge the technical support of Dan Coughlin, Jamie Marsh and Yi Yi, all from the University of Georgia's Savannah River Ecology Laboratory. Most of the funding for this research was provided from The Environmental Restoration Division of Westinghouse Savannah River Company. Partial funding was provided through Financial Assistance Award Number DE-FC09-96SR18546 from the U.S. Department of Energy to the University of Georgia Research Foundation.

7.0 REFERENCES

Abraham, J.P., F.W. Whicker, T.G. Hinton, and D.J. Rowan. 2000. "Inventory and spatial pattern of ¹³⁷Cs in a pond: a comparison of two survey methods." *Journal of Environmental Radioactivity* 51:157-171.

Archer, F.C. 1971. Factors affecting the trace element content of pastures. In: Trace elements in soils and crops. *Min Ag Fish Fd Tech Bull, HMSO*, 21:150-157.

Frnacis, C.W., and F.S. Brinkely. 1976. "Preferential adsorption of ¹³⁷Cs to micaceous minerals in contaminated freshwater sediment." *Nature* 260: 511-513.

Garten, Jr., C.T., and D. Paine. 1977. "A multivariate analysis of factors affecting radiocesium uptake by *Sagittaria latifilia* in coastal plain environments." *Journal of Environmental Quality* 6(1):78-82.

Hall, G.E.M., JE. Vaive, R. Beer, and Hoashi. 1995. "Selective leaches revisited, with emphasis on the amorphous Fe oxyhydroxide phase extraction." *Journal of Geochemical Exploration* 56:59-78.

Hinton, T.G., C..M. Bell, F.W. Whicker, and T. Philippi. 1999. "Temporal changes and factors influencing ¹³⁷Cs concentration in vegetation colonizing an exposed lake bed over a three-year period." *Journal of Environmental Radioactivity* 44:1-19.

Horrill, A.D., V.H. Kennedy, and T.R. Harwood. 1990. The concentrations of Chernobyl derived radionuclides in species characteristic of natural and semi-natural ecosystems. In Proceedings of the Workshop Transfer of Radionuclides in Natural and Semi-Natural Environments (pp.27-39). Passarino, Italy, 11-15 September 1989.

Hughes, M.K., N.W. Lepp, D.A. Phipps. 1980. "Aerial heavy metal pollution and terrestrial ecosystems." *Adv. Ecol. Res.* 11:217-327.

Kaplan, D. I., A. S. Knox, T. G. Hinton, R. R. Sharitz, B. P. Allen, and S. M. Serkiz. 2001. Proof-of-Concept of the Phytoimmobilization Technology for TNX Outfall Delta WSRC-TR-2001-00032, Westinghouse Savannah River Company, Aiken, South Carolina.

Miller, W.P., D.C. Martens, L.W. Zelazny, and E.T. Kornegay. 1986. "Forms of Solid Phase copper in Copper-enriched Swine Manure." Journal of Environmental Quality 15:69-72.

Miller, W.P. and D.M. Miller. 1987. "A Micro-Pipette Method for Soil Mechanical Analysis." *Communications In Soil Sci.*, *Plant Anal.* 18(1) 1-15.

NCRP, National Council on Radiological Protection and Measurements. 1989. Screening techniques for determining compliance with environmental standards. NCRP Commentary No. 3. Bethesda, Maryland.

Pinder, J.E. III, C.T. Garten, and D. Paine.1980. "Factors affecting radiocesium uptake by plants inhabiting a contaminated flood plain." *Acta Oecologica* 1:3-10.

Seel, J.F., F.W. Whicker, and D.C. Adriano. 1995. "Uptake of ¹³⁷Cs in vegetable crops grown on a contaminated lakebed." *Health Physic* 68(6):793-799.

Thomas, G.W. 1996. Soil pH and Soil Acidity. In: D.H. Sparks (ed.) Methods of Soil Analysis, Part 3, Chemical Methods, Soil Science Society of America and American Society of Agronomy, Madison, Wisconsin.

Tessier, A., P.G.C. Campbell, and M. Bisson. 1979. "Sequential extraction procedure for the speciation of particular trace metals." *Analytical Chemistry*. 51:844851.

Whicker, F.W., and V. Schultz. 1982. Radioecology: Nuclear Energy and the Environment. CRC Press, Boca Raton, Florida.